

Epidemiology and antibiogram of Coagulase Negative Staphylococci (CoNS) Isolated from various Clinical Samples in Tertiary Care Teaching Hospital in South India

¹S. Nagaraju, Tutor , Department of microbiology , Mamata Academy of Medical Sciences, Bachupally, Hyderabad - 500090

²B. Harikrishna, Research Scientist, Medal Health Care Pvt. Ltd, Karnool-518002

Corresponding Author: S. Nagaraju, Tutor, Department of Microbiology, Mamata Academy of Medical Sciences, Bachupally, Hyderabad - 500090

Type of Publication: Original Research Article

Conflicts of Interest: Nil

Abstract

Introduction and Objectives: In the last two decades CoNS has become recognized as a leading cause of nosocomial bacteremia, wound, bone, joint and urinary tract infections (UTI's). CoNS are one of the most frequent causes of nosocomial infections and are reservoirs of multiple antimicrobial resistant determinants. Hence, this study was done to appreciate and to determine the sensitivity pattern of CoNS.

Materials and Methods: The present study will be carried out from April to June, 2018, where in 490 CoNS isolated from various clinical samples like urine, pus, body fluids, blood, sputum, and catheter tips etc & submitted to the department of microbiology in SVIMS Hospital, Tirupathi.

Results: Out of 490 CoNS species isolated majority were from blood samples 52%, rest were from pus 23%, urine 8%, catheter tip 7%, body fluids 6% & others 4%. Among 490 CoNS, majority were *S.hominis* (47%) and *S.hemolyticus* (41%) followed by other species. Methicillin Resistance was noted high in *S.epidermidis* (60%) followed by *S.hemolyticus* (32%), *S.saprophyticus* (20%), *S.schleiferi* (20%) & *S.hominis* (16%).

Conclusion: The increased recognition of pathogenic potential in CoNS and emergence of drug resistance

among them shows the necessity to adopt simple lab methods to identify the species and determine the antibiotic resistant patterns which will be helpful in effective management of patients infected with CoNS.

Keywords: Coagulase negative Staphylococcus, Antimicrobial susceptibility pattern, Methicillin resistance.

Introduction

The Coagulase Negative Staphylococcus (CoNS) is a significant nosocomial pathogen has increased in recent years. Enormous increase and emergence of these strains has simultaneously led to a variety of infections and these organisms are also resistance to various antimicrobial agents. Nosocomial infections due to CoNS has increased and there found reservoirs of multiple antimicrobial resistant determinants [1-3]. CoNS, formerly regarded as harmless inhabitants of the skin and mucosal linings, may be present in a specimen as a contaminant or a pathogen [4-6]. CoNS are the commonly encountered pathogens in nosocomial infections, partly due to the growing appreciation of these groups of organisms as opportunistic pathogens or due to increase in the use of medical devices such as catheters, shunts, heart valve graft material, implants and artificial joints in seriously debilitating and immune-compromised patients [7]. Drug users are at increased risk of infections due to CoNS [3].

More than 30 species of CoNS are recognized but only a few are commonly incriminated in human infections [8]. Most frequently isolated species of CoNS from infections is *S.epidermidis*. In the reports of national survey, this species has been remarkably quoted as primary nosocomial pathogen [9]. It has been implicated as the etiological agent in infections of wounds, urogenital tract, respiratory tract, meninges, conjunctiva and intravenous catheter associated infections [10]. *S.haemolyticus* has been documented as the second most common CoNS species recovered from documented infection sites. It has been implicated in naïve-valve endocarditis, septicaemia, peritonitis, wound, bone and joint infections [11]. *S. warneri* is a well recognised cause of catheter related bacteraemia, naïve-valve endocarditis, haematogenous vertebral osteomyelitis and ventriculo-peritoneal shunt associated meningitis [12]. *S.saprophyticus* was shown to be an important cause of urinary tract infections in young females. It is second to coliforms as the most common cause of acute urethral syndrome [13 & 14]. *S.hominis* has occasionally been isolated from infections causing catheter related sepsis in immune-compromised hosts [15]. *S.simulans* has been established as a cause of septicemia, osteomyelitis, septic arthritis, vertebral osteomyelitis and prosthetic joint infection [16]. *S.schleiferi* has been isolated from several human infections including brain empyema, wound infections, bacteraemia complicating vertebral osteitis, infection of hip prosthesis and indwelling catheter infections [17]. *S.lugdunensis* has been isolated from abscesses in the pelvic girdle region [18]. *S.cohnii* is an emergent opportunistic agent having been reported as a cause of community acquired pneumonia [19].

The postulated reasons for the current prevalence and clinical importance of these organisms include their great number on the skin, their selection as a result of wide

spread usage of broad spectrum antibiotics in the hospitals, their ability to adhere and form bio- films on the surface of vascular catheters and other medical devices [2].

The present study was undertaken, to identify the most prevalent clinical isolates of CoNS by minimum number of tests necessary and sufficient to discriminate between the species & to determine the antimicrobial susceptibility pattern of these isolates.

Materials and Methods

The present study will be carried out from April to June, 2018 in the Department of Microbiology, Sri Venkateswara institute of Medical Sciences, Tirupathi. CoNS will be isolated from various clinical samples like urine, pus, body fluids, blood, sputum, and catheter tips etc submitted to the department of microbiology in SVIMS Hospital, Tirupathi.

The samples will be inoculated on Nutrient agar, Blood agar and Mac Conkey agar. The inoculated plates will be incubated at 37⁰C overnight. In case of any growth on the plates, it will be processed according to the standard bacteriological techniques. The colonial appearance and morphological characteristics of the isolated bacteria will be noted. The isolated colonies will be subjected to preliminary tests like Gram staining, Catalase test, Coagulase (slide & tube) test and sugar fermentation tests. These preliminary tests will be followed by various biochemical tests to speciate CoNS as follows

1. Coagulase test (Slide & Tube Coagulase tests)
2. Urease test.
3. Sugar fermentation test (xylose, arabinose, sucrose, trehalose, maltose, mannitol).
4. Susceptibility to Polymixin-B and Novobiocin.
5. Anaerobic growth in thioglycollate broth

Results

A total of 490 CoNS isolates were obtained from various clinical samples. Out of 490 CoNS isolated, 51% were from male and 49% from female patients.(Fig:1)

52% were isolated from blood samples, rest from pus (23%), urine (8%), catheter tips (7%), body fluids (6%), others (4%). (Fig:2).

Among 490 CoNS, majority were S.hominis (47%) and S.hemolyticus (41%) followed by S.epidermidis (8%), S.saprophyticus (2%), S.schleiferi (2%). (Fig:3)

Methicillin Resistance was noted high in S.epidermidis (60%) followed by S.hemolyticus (32%), S.saprophyticus (20%), S.schleiferi (20%) & S.hominis (16%). (Fig:4)

Resistance to Vancomycin which is the drug of choice for methicillin resistant strains was observed in all the species of CoNS in the present study, which is an alarming sign. (Table:1)

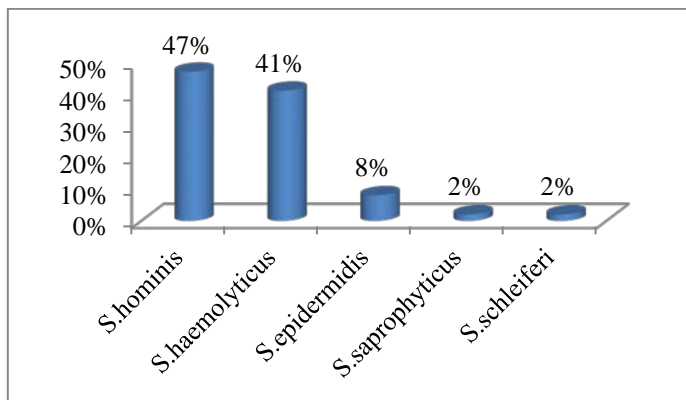


Fig. 3: Species wise distribution of CoNS

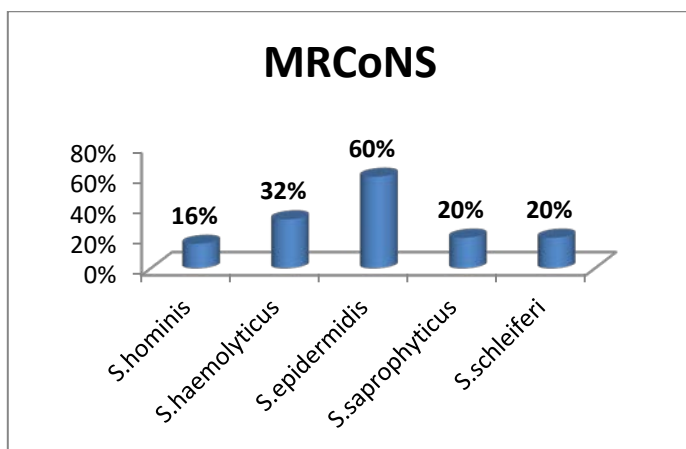


Fig. 4: Distribution of MRCoNS among isolated species

Table 1: Antibiotic sensitivity pattern of isolated CoNS species

Sn	Organism	CFN	CF	CD	ER	CO T	LZ	VA N
1	S.hominis (229)	16%	24 %	26 %	36 %	41%	2 %	Nil
2	S.haemolyticus (203)	32%	38 %	28 %	41 %	41%	5 %	Nil
3	S.epidermidis (38)	60%	56 %	68 %	64 %	79%	6 %	Nil
4	S.saprophyticus (10)	20%	Nil	Nil	Nil	10%	Ni 1	Nil
5	S.schleiferi f(10)	20%	Nil	10 %	10 %	20%	Ni 1	Nil

Legends: CFN- Cefoxitin, CF-Ciprofloxacin, CD-Clindamycin, ER-Erythromycin, COT Cotrimaxazole, LZ-Linezolid, VAN-Vancomycin
Legends: CoNS- Coagulase negative staphylococcus

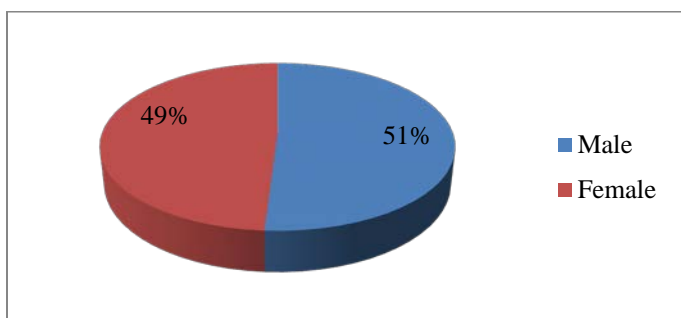


Fig. 1: Gender wise distribution of CoNS

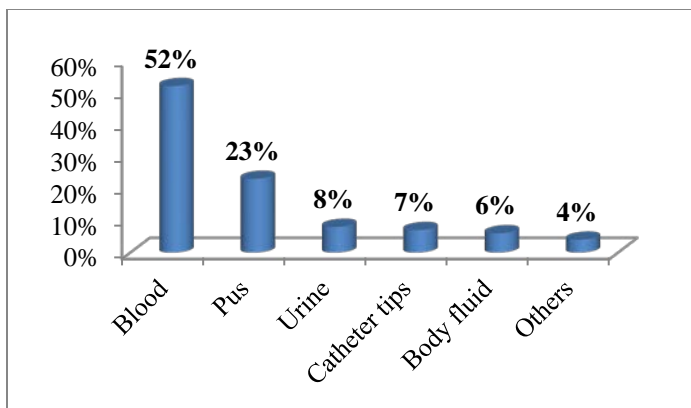


Fig. 2: Sample wise distribution of CoNS

MRCoNS- Methicillin resistant coagulase negative staphylococcus

Discussion

CoNS are generally considered normal inhabitants of skin and nares. CoNS are capable of causing only opportunistic infections, so many clinical laboratories do not identify clinical isolates of CoNS to the species level. As CoNS is increasingly being implicated as significant nosocomial pathogens, several reviewers has emphasized the need for species identification, which is possible only by a simple, easily adaptable, inexpensive method [4,5]. The species identification is important in monitoring the reservoir and distribution of CoNS involved in nosocomial infections and determining the etiological agent [6]. In our study *S.hominis* (47%) was the most common isolate. This correlates with other studies by K. Punitha Valli et al. (32.3%) [20], Saroj Golia et al. (46.3%) [21], C. Roopa et al. (50%) [22], S.S. Vijayasri Badampudi et al. (40%) [23]. The remaining species isolated were *S.haemolyticus* (41%), *S.saprophyticus* (2%), and *S.Epidemidis*(8%) *S.schleiferi* (2%). This prevalence is in concordance with various other studies. [20-23].

Conclusion

In our study most frequently encountered clinical isolates in our hospital were *S.hominis* (47%), followed by *S.haemolyticus* (41%), *S.epidemidis* (8%), *S.saprophyticus* (2%), *S.schleiferi* (2%). The present study revealed maximum number of CoNS isolates from blood samples. Differences in antibiotic sensitivity patterns indicate that speciation of CoNS is most useful for clinical and epidemiologic purposes. High resistance to antimicrobials was observed among *S.epidermidis* and *S.hemolyticus* which is alarming. So regular reporting & monitoring of antimicrobial resistance patterns of CoNS species is imperative to correlate clinically.

References

1. Goyal R, Singh NP, Kumar A, Kaur I, Singh M, Sunita N, Mathur M. Simple and economical method for speciation and resistotyping of clinically significant coagulase negative staphylococci. *Indian J Med Microbiol.* 2006 Jul;24(3):201-4.
2. Pfaller MA, Herwaldt LA. Laboratory, clinical, and epidemiological aspects of coagulase-negative staphylococci. *Clinical Reviews.* 1988;1(3):281-299.
3. Longauerova A. Coagulase-negative staphylococci and their participation in pathogenesis. *Bratisl Lek Listy* 2006; 107(11-12):448-452.
4. Washington Winn JR, Stephen Allen, William Janda, Elmer Koneman, Gary procop, Paul Schreckenberger, Gail Woods, editors. Lippincott William & Wilkins; *Staphylococci and related gram-positive cocci. Gram-positive cocci,chapter-12. In: konemanns color Atlas and text book of DaignosticMicrobiology, Sixth edition.* 2006. P. 624-671.
5. Mohan U, Jindal N, Aggarwal P. Species distribution and antibiotic sensitivity pattern of coagulase-negative staphylococci isolated from various clinical specimans. *Indian J Med Microbiol* 2002; 20:45-46.
6. Cunha, Mlrs, Sinzato, YK and Silveira LVA (2004) comparison of methods for the identification of coagulase- negative staphylococci. *Memorials do Instituto Oswaldo Cruz.* 99, 855-860.
7. Badwi JA, Memon AH, Soomro AA. Coagulase negative *Staphylococcus* (CONS) is the contaminant in the clinical specimen. *Med Channel* 2012;19:23-7.
8. Geary C, Jordens JZ, Richardson JF, Howcraft DM, Mitchell CJ. Epidemiological typing of Coagulase negative *Staphylococci* from nosocomial infections. *J Med Microbiol* 1997;46:195-203.
9. Fule RP, Later Iyer, Saoji AM. Study of pathogenicity markers of *Staphylococci* isolated from clinical

- specimens. *Indian J PatholMicrobiol* 1996;39(2):127-130
10. Joshi JR, Pawar S, Joshi PJ, Samuel A. Biological characters and sensitivity of *Staphylococcus epidermidis* *Indian J PatholMicrobiol* 1987;30:89-96.
11. Schwalbe RS, Ritz WJ, Verma PR, Barranco EA, Gilligan PH. Selection for vancomycin resistance in clinical isolates of *Staphylococcus haemolyticus*. *The J Infect Dis* 1990; 161:45-51
12. Picket DA, Welch DF. Recognition of *Staphylococcus saprophyticus* in urine cultures by screening colonies for production of phosphatase. *J ClinMicrobiol* 1985;21:310-313.
13. Marrie TJ, Kwan C, Noble MA, West A, Duffield L. *Staphylococcus saprophyticus* as a cause of urinary tract infections. *J ClinMicrobiol* 1982;16(3):427-431.
14. Picket DA, Welch DF. Recognition of *Staphylococcus saprophyticus* in urine cultures by screening colonies for production of phosphatase. *J ClinMicrobiol* 1985;21:310-313.
15. Bannerman TL. *Staphylococcus*, *Micrococcus* and other catalase positive cocci that grows aerobically Chapter 28. In: *Manual of ClinicalMicrobiology*, 8th ed. Murray PR, Baron EJ, Jorgensen JH, Pfaller MA, Tenover FC, Tenover FC, editors. ASM press: Washington DC; 2003 p.384.
16. De paulis AN, Predari SC. Five test simple scheme for the species level identification of clinically significant coagulase negative staphylococci. *J ClinMicrobiol*. 2003; 41: 1219-1224
17. Surekha Y Asangi, Mariraj J, Satyanarayana M S, Nagabhushan, Rashmi. Speciation of clinically significant Coagulase Negative Staphylococci and their antibiotic resistant patterns in a tertiary care hospital. *Int J Biol Med Res*. 2011;2(3):735-739.
18. Shubhra Singh, Gopa Banerjee et al. Prevalence of *MecA* Gene positive coagulase negative Staphylococci in NICU of a tertiary care hospital. *Biomedical Research*. 2009; 20 (2): 94-98.
19. Sheikh AF, Mehdinejad M. Identification and determination of coagulase negative Staphylococci species and antimicrobial susceptibility pattern of isolates from clinical specimens. *Afr J Microbiol Res* 2012;6:1669-74.
20. Valli KP, Pramodhini S, Umadevi S, Seetha KS. Speciation and Detection of Virulence Factors of Coagulase Negative Staphylococci Isolated from Various Clinical Samples. *Int. J. Curr. Microbiol. App. Sci*. 2016;5(4):159-64.
21. Golia S, Telsang DB, Kamath BA, Tiwari D. Speciation of clinically significant coagulase negative staphylococci and their antibiotic resistant patterns in a tertiary care hospital.
22. Roopa C, Biradar S. Incidence and speciation of coagulase negative staphylococcus isolates from clinically relevant specimens with their antibiotic susceptibility patterns. *Int. J. Curr. Microbiol. Appl. Sci*. 2015;4(9):975-80.
23. Badampudi SS, KRL SK, Gunti R. Speciation and Biofilm Production of Coagulase Negative Staphylococcal Isolates from Clinically Significant Specimens and their AntibioGram. *Journal of Krishna Institute of Medical Sciences (JKIMSU)*. 2016 Apr 1;5(2).

How to citation this article: S. Nagaraju, B. Harikrishna, "Epidemiology and antibiogram of Coagulase Negative Staphylococci (CoNS) Isolated from various Clinical Samples in Tertiary Care Teaching Hospital in South India", *IJMACR*- November - December - 2020, Vol – 3, Issue -6, P. No. 116 – 121.

Copyright: © 2020, S. Nagaraju, et al. This is an open access journal and article distributed under the terms of the creative commons attribution noncommercial License 4.0. Which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.
